



The central nervous system is not immunoprivileged: inflammation and epileptogenesis

Centralni nervni sistem nije imunoprivilegovan: inflamacija i epileptogeneza

Dragan Hrnčić*, Nikola Šutulović*, Željko Grubač*,
Aleksandra Rašić-Marković*, Olivera Stanojlović*

University of Belgrade, Faculty of Medicine, Institute of Medical Physiology “Richard Burian”, *Laboratory of Neurophysiology, Belgrade, Serbia

Key words:
central nervous system; epilepsy; inflammation;
oxidative stress.

Ključne reči:
nervni sistem, centralni; epilepsija; zapaljenje; stres,
oksidativni.

Introduction

Epilepsy is a chronic neurological disorder characterized by seizures which are the result of excitation/inhibition disbalance of neurotransmitters in the central nervous system (CNS)¹. The main substrate in the occurrence of seizures is a transient, sudden, paroxysmal and hypersynchronous activity of the brain neurons, behaviorally manifested by rapid and repetitive skeletal muscle contractions and relaxations². A process of gradual epilepsy development in previously healthy brain is known as epileptogenesis³.

Approximately 0.5%–1% (about 65 million) of adults and 0.5% of children suffer from epilepsy in the world, while 2 out of 25 people in the general world population had at least one seizure in their lifetime⁴. These facts reflect the impact of epilepsy on the social, economic and emotional aspects of life of these patients as well as treatment costs of this neurological disorder with multiple sociomedical consequences. It is estimated that 6 million people suffer from epilepsy in Europe and 50 thousand people in Serbia nowadays. More than 65% of patients experienced a first seizure attack in childhood (the highest incidence is in the first years of life). Finally, there is another important fact: about 40% of patients with epilepsy are resistant to current antiepileptic therapy⁵.

The central nervous system (CNS) homeostasis is dependent on the balance of two opposite processes in the brain, e.g., excitation and inhibition. Thus, even minimal disequilibrium between these two processes (i.e., increase of the excitation, decrease of the inhibition, or both) will lead to hyperexcitability and will cause seizures⁶. Molecular pathways

of epileptogenesis are still an enigma, and it is consequently difficult to establish the precise classification of epileptic disorders and prevent seizures completely (despite significant progress in the new antiepileptic drugs development).

Epilepsy, as a chronic disorder, is characterized by spontaneous and recurrent seizures caused by certain pathogenic processes that disrupt neuronal and glial cells structure and/or function. Factors that cause epilepsy and provoke individual, isolated convulsions are numerous and very diverse. The most common among them are: fever in children, tricyclic antidepressants, theophylline, drugs intoxications and drugs abuse, acute neurological disorders and infections (meningitis, encephalitis, stroke, head injury, brain abscess, etc.), alcoholism, metabolic disorders (hypo/hyperglycemia, hypocalcaemia)⁷, derivative of phencyclidine-metaphit⁸, scabicides-lindane^{9, 10}, sleep disturbances^{11–13}, hyperhomocysteinemia¹⁴, visual stimuli¹⁵, stress¹⁶, the menstrual cycle¹⁷, a specific diet regimes^{18, 19}, vascular abnormalities, stroke sequelae and subarachnoid hemorrhage²⁰. Experimental models of epilepsy are significant for resolving the mechanisms of epileptogenesis and play very important role in new antiepileptic drugs development. Extensive review of all models of epilepsy goes beyond 50 entries²¹.

Some antibiotics, like imipenem/cilastatin^{22, 23} and penicillin²⁴ induce seizures which is also a paradoxical phenomenon clearly demonstrated in experimental and clinical studies.

Inflammation, engaging the immune cells, the molecular mediators and blood vessels, is a complex biological, protective response to the pathogenic factors²⁵. In conditions of excessive activity and/or organism immunity collapse, in-

flammation seems to be potentially harmful and unuseful. Neuroinflammation could be caused by numerous factors and is characterized by activation of glial cells, known as the CNS resident immune cells²⁶. Blood-brain barrier (BBB) is built up of astrocytes and endothelial cells and acts as an important isolator of the CNS. Therefore, the CNS is also partially isolated from the peripheral inflammatory cells²⁷. Peripheral inflammation is an inflammatory process which occurs outside the CNS and is characterized by the activation of macrophages (peripheral immune cells)²⁸. When the BBB is disrupted or its function compromised, peripheral immune cells could migrate into the CNS and induce neuroinflammation²⁹.

Inflammatory processes and epileptogenesis

Numerous epidemiological studies have demonstrated that patients with epilepsy are more prone to different inflammatory disorders, i.e., inflammatory diseases are frequent comorbidities in epilepsy³⁰.

Numerous clinical and experimental studies have shown that inflammation disrupts the excitation/inhibition balance within the CNS and initiates the process of epileptogenesis. However, unclear relationship exists among the convulsions, inflammation and epilepsy^{31–36}. These three factors generate positive feedback loop (*circulus vitiosus*) in which one process is facilitated by another one (Figure 1). Head injuries, tumors and infections of the CNS are characterized by neuroinflammation in its pathological basis. On the other hand, neuroinflammation could be a result of these pathological processes. Also, it is important to notice that seizure could be followed by inflammatory response in the entire CNS³⁷. The CNS is not immunoprivileged site in the human body.

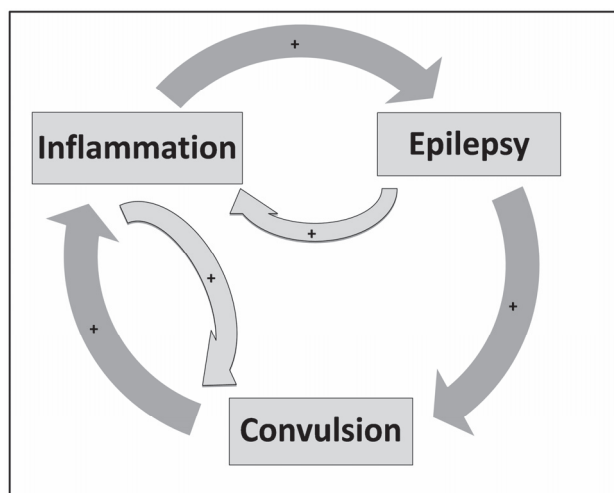


Fig. 1 – The relationship between epilepsy, convulsions and inflammation.

Neuroinflammation and peripheral inflammation could be a etiopathogenic substrate of epileptogenesis and development of epilepsy as a chronic neurological disorder accompanied by convulsive seizures. On the other hand, inflammation could also destroy the excitation/inhibition balance in the central nervous system (CNS) and provoke only isolated convulsions. On the other hand, isolated convulsive seizure could provoke neuroinflammation. Thus, positive feedback loop exists among these factors.

The process of epileptogenesis should not be viewed only through the prism of neural networks disarrangement. Namely, the pathogenesis of epilepsy involves also disarray of non-neural structures such as glial cells, endothelial cells in the CNS, the peripheral leukocytes and BBB injury³⁸. Also, the role of extracellular matrix (ECM), as a very important brain substrate consisting of various molecules [e.g., hyaluronan, chondroitin sulfate proteoglycans (CSPGs), glycoprotein tenascin-R and other] derived from both neurons and glial cells³⁹ has to be addressed in these considerations, having in mind a critical role of cell-ECM interactions. Namely, there is a bidirectional relationship between epileptogenesis and ECM as well as evidences showing that biophysical and biochemical ECM properties modulate immune cell behavior including inflammatory cell migration. Excessive neuronal activity-mediated remodeling of ECM and mutations in ECM molecules (like leucine-rich, glioma-inactivated gene (LGI1), hyaluronan, tissue plasminogen activator (tPA) etc.) and extracellular proteinases [i.e. matrix metalloproteinase-9 (MMP-9)] are potent triggers for epileptogenesis by still unclear mechanisms which include disbalances in GABAergic and glutamatergic activity, mossy fiber sprouting, granule cell dispersion and gliosis (further details could be find in recent focused reviews^{39–41}). During inflammation, ECM is altered by several cytokines upregulated at sites of inflammation [like tumor necrosis factor (TNF) and interferon gamma (IFN γ)] as well as by the secretion and/or activation of proteases, like MMPs⁴². On the other hand, neuroinflammation leads to degradation of ECM into immunoreactive fragments (mainly hyaluronan, sulfated proteoglycans and newly expressed tenascin-C), which are considered as damage-associated molecular patterns (DAMPs) capable to activate inflammatory cells via pattern recognition receptors (PRRs)⁴³. Hence, progression of neuroinflammation, in turn, leads to further degradation of ECM by positive feedback loop. Therefore, ECM protects neuronal cell rearrangement and excitatory network formation by its nonpermissive function, and any change in neural ECM induced by inflammation, brings a receptive environment for epileptogenesis.

The immune response is proven to be responsible for the initiation and propagation of epilepsy^{37, 44, 45}. Neuroinflammation increases the excitability of neurons because it changes the permeability of neurilemmal ion channels, affects the release and reuptake of neurotransmitters and the permeability of the BBB. All these factors decrease the threshold for the occurrence of epileptic discharges^{46–48}.

Oxidative stress and neuroinflammation: unresolved relationship

Neuroinflammation occurs by activation of glial cells (astrocytes, ependymal and endothelial cells in the first row) with the increased production of proinflammatory cytokines [like interleukin-1 β (IL-1 β); tumor necrosis factor- α (TNF- α); prostaglandins, etc.] at the same time. It is also followed by an increased activity of cyclooxygenase-2 (COX-2)³⁷. IL-1 receptor/toll-like receptor (IL-1R/TLR) and COX-2-medi-

Peripheral inflammation is accompanied by an increased production of proinflammatory cytokines, like IL-1 β , IL-6 and TNF- α , which may contribute to the activation of glial cells and induction of neuroinflammation⁶⁶⁻⁶⁹. On the other hand, the role of COX-2 was emphasized in the process of neuroinflammation and oxidative stress, both acting to reduce the seizure threshold^{70, 71}. The concentration and activity of the COX-2 enzyme are markedly increased during seizures⁷², especially in the hippocampus⁷³ which has essential role in the epilepsy generalization. Indeed, Ho et al.⁷⁰ showed reduction of seizure number in the experimental model of peripheral inflammation accompanied by neuroinflammation and oxidative stress in the hippocampus upon intracerebral administration of a COX-2 inhibitor and ROS scavenger. Also Eun et al.⁷⁴ potentiated hyperthermia-induced seizures by administration of LPS, which is a further proof that peripheral inflammation can potentiate seizures.

Conclusion

Many neurological conditions and disorders, in the first row inflammatory diseases of the CNS (meningitis, encephalitis), CNS injuries, stroke, etc., are associated with inflam-

mation. On the other hand, clinical course of these disorders frequently includes seizures. Inflammation is a common factor that disrupts homeostasis in the CNS neural networks; it acts proepileptogenically and causes seizures.

Taking into account all the above-mentioned facts, it is clear that neuroinflammation, together with its specific relation to ROS and changes in neural ECM, is interconnected with epileptogenesis, being its cause, or a consequence. Neuroinflammation reduces the threshold for initiation of the CNS hyperexcitability and disrupts the competency of the BBB.

It is reasonable to assume that therapeutic treatment of peripheral inflammation alongside with neuroinflammation can provide beneficial effects for patients with epilepsy by reducing the seizure number. Thus, this is a promising alley of future therapeutic modalities in epileptology.

Acknowledgement

This work was supported by the Ministry of Education, Science and Technological Development of Serbia (Grant No175032).

R E F E R E N C E S

1. Riazi K, Galic MA, Pittman QJ. Contributions of peripheral inflammation to seizure susceptibility: Cytokines and brain excitability. *Epilepsy Res* 2010; 89(1): 34-42.
2. Fisher RS, van Emde BW, Blume W, Elger C, Genton P, Lee P, et al. Epileptic seizures and epilepsy: Definitions proposed by the International League Against Epilepsy (ILAE) and the International Bureau for Epilepsy (IBE). *Epilepsia* 2005; 46(4): 470-2.
3. McNamara JO, Huang YZ, Leonard AS. Molecular signaling mechanisms underlying epileptogenesis. *Sci STKE* 2006; 2006(356): re12.
4. World Health Organization. Epilepsy. Epilepsy: Epidemiology, aetiology and prognosis. Fact Sheet No.165. Geneva: World Health Organization; 2001.
5. World Health Organization. Epilepsy. Epilepsy: Epidemiology, aetiology and prognosis. Fact Sheet Update. Geneva: World Health Organization; 2016.
6. Heise C, Taba E, Murru L, Ponzoni L, Cattaneo A, Guarnieri FC, et al. eEF2K/eEF2 Pathway Controls the Excitation/Inhibition Balance and Susceptibility to Epileptic Seizures. *Cereb Cortex* 2017; 27(3): 2226-48.
7. Shneker BF, Fountain NB. Epilepsy. *Dis Mon* 2003; 49(7): 426-78.
8. Stanojlović O, Živanović D, Šušić V. N-Methyl-D-aspartic acid- and metaphit-induced audiogenic seizures in rat model of seizures. *Pharmacol. Res* 2000; 42(3): 247-53.
9. Hrnčić D, Rašić-Marković A, Đurić D, Sušić V, Stanojlović O. The role of nitric oxide in convulsions induced by lindane in rats. *Food Chem Toxicol* 2011; 49(4): 947-54.
10. Mladenović D, Hrnčić D, Vučević D, Radosavljević T, Lončar-Stevanović H, Petrović J, et al. Ethanol suppressed seizures in lindane-treated rats. *Electroencephalographic and behavioral studies. J Physiol Pharmacol* 2007; 58(4): 641-56.
11. Šušić V, Marković O. Potentiation of metaphit-induced audiogenic seizures by REM sleep deprivation in rats. *Physiol Behav* 1993; 54(2): 331-8.
12. Hrnčić D, Rašić-Marković A, Bjekić-Macut J, Šušić V, Đurić D, Stanojlović O. Paradoxical sleep deprivation potentiates epilepsy induced by homocysteine thiolactone in adult rats. *Exp Biol Med (Maywood)* 2013; 238(1): 77-83.
13. Hrnčić D, Grubač Ž, Rašić-Marković A, Šutulović N, Šušić V, Bjekić-Macut J, et al. Sleep disruption increases seizure susceptibility: Behavioral and EEG evaluation of an experimental model of sleep apnea. *Physiol Behav* 2016; 155: 188-94.
14. Stanojlović O, Rašić-Marković A, Hrnčić D, Šušić V, Macut D, Radosavljević T, et al. Two types of seizures in homocysteine thiolactone-treated adult rats, behavioral and electroencephalographic study. *Cell Mol Neurobiol* 2009; 29(3): 329-39.
15. Zifkin BG, Inoue Y. Visual Reflex Seizures Induced by Complex Stimuli. *Epilepsia* 2004; 45(Suppl 1): 27-9.
16. Moon HJ, Seo JG, Park SP. Perceived stress and its predictors in people with epilepsy. *Epilepsy Behav* 2016; 62: 47-52.
17. Herzog AG, Fowler KM, Sperling MR, Massaro JM. Progesterone Trial Study Group. Distribution of seizures across the menstrual cycle in women with epilepsy. *Epilepsia* 2015; 56(5): e58-62.
18. Gordon KE, Dooley JM. Food insecurity and epilepsy in a nationally representative sample. *Epilepsy Behav* 2015; 43: 139-42.
19. Hrnčić D, Rašić-Marković A, Stojković T, Velimirović M, Puškaš N, Obrenović R, et al. Hyperhomocysteinemia induced by methionine dietary nutritional overload modulates acetylcholinesterase activity in the rat brain. *Mol Cell Biochem* 2014; 396(1-2): 99-105.
20. Annegers JF, Rocca WA, Hauser WA. Causes of epilepsy: Contributions of the Rochester epidemiology project. *Mayo Clin Proc* 1996; 71(6): 570-5.
21. Stanojlović O, Živanović D. Experimental models of epilepsy. *Med Pregl* 2004; 57(7-8): 359-62.

22. Živanović D, Stanojlović OS, Šušić V. Effects of manipulation of N-methyl-D-aspartate receptors on imipenem/cilastatin-induced seizures in rats. *Indian J Med Res* 2004; 119(2): 79–85.
23. Živanović D, Stanojlović O, Stojanović J, Šušić V. Induction of audiogenic seizures in imipenem/cilastatin-treated rats. *Epilepsy Behav* 2004; 5(2): 151–8.
24. Tokiwa T, Inoue T, Fujii M, Ishizuka S, Aou S, Kida H, et al. Penicillin-induced epileptiform activity elevates focal brain temperature in anesthetized rats. *Neurosci Res* 2013; 76(4): 257–60.
25. Ferrero-Miliani L, Nielsen OH, Andersen PS, Girardin SE. Chronic inflammation: Importance of NOD2 and NALP3 in interleukin-1beta generation. *Clin Exp Immunol* 2007; 147(2): 227–35.
26. Gendelman HE. Neural immunity: Friend or foe? *J Neurovirol* 2002; 8(6): 474–9.
27. Das Sarma J. Microglia-mediated neuroinflammation is an amplifier of virus-induced neuropathology. *J Neurovirol* 2014; 20(2): 122–36.
28. di Filippo M, Chiasserini D, Gardoni F, Viviani B, Tozzi A, Giampà C, et al. Effects of central and peripheral inflammation on hippocampal synaptic plasticity. *Neurobiol Dis* 2013; 52: 229–36.
29. 't Hart BA, den Dunnen WF. Commentary on special issue: CNS diseases and the immune system. *J Neuroimmune Pharmacol* 2013; 8(4): 757–9.
30. Téllez-Zenteno JF, Matijević S, Wiebe S. Somatic comorbidity of epilepsy in the general population in Canada. *Epilepsia* 2005; 46(12): 1955–62.
31. Majores M, Eils J, Wiestler OD, Becker AJ. Molecular profiling of temporal lobe epilepsy: Comparison of data from human tissue samples and animal models. *Epilepsy Res* 2004; 60(2–3): 173–8.
32. Ravizza T, Gagliardi B, Noé F, Boer K, Aronica E, Vezzani A. Innate and adaptive immunity during epileptogenesis and spontaneous seizures: Evidence from experimental models and human temporal lobe epilepsy. *Neurobiol Dis* 2008; 29(1): 142–60.
33. Aronica E, Crino PB. Inflammation in epilepsy: Clinical observations. *Epilepsia* 2011; 52(Suppl 3): 26–32.
34. Janigro D, Tffland PH, Marchi N, Granata T. A role for inflammation in status epilepticus is revealed by a review of current therapeutic approaches. *Epilepsia* 2013; 54(Suppl 6): 30–2.
35. Godukhin OV, Levin SG, Parnyskhova YE. The effects of interleukin-10 on the development of epileptiform activity in the hippocampus induced by transient hypoxia, bicuculline, and electrical kindling. *Neurosci Behav Physiol* 2009; 39(7): 625–31.
36. Kawamura Y, Yamazaki Y, Ohashi M, Ihira M, Yoshikawa T. Cytokine and chemokine responses in the blood and cerebrospinal fluid of patients with human herpesvirus 6B-associated acute encephalopathy with biphasic seizures and late reduced diffusion. *J Med Virol* 2014; 86(3): 512–8.
37. Vezzani A, Granata T. Brain inflammation in epilepsy: Experimental and clinical evidence. *Epilepsia* 2005; 46(11): 1724–43.
38. Xu D, Miller SD, Koh S. Immune mechanisms in epileptogenesis. *Front Cell Neurosci* 2013; 7: 195.
39. Dityatev A. Remodeling of extracellular matrix and epileptogenesis. *Epilepsia* 2010; 51(Suppl 3): 61–5.
40. Dityatev A, Fellin T. Extracellular matrix in plasticity and epileptogenesis. *Neuron Glia Biol* 2008; 4(3): 235–47.
41. Pitkänen A, Ndoe-Ekane XE, Łukasiuk K, Wilczynski GM, Dityatev A, Walker MC, et al. Neural ECM and epilepsy. *Prog Brain Res* 2014; 214: 229–62.
42. Sorokin L. The impact of the extracellular matrix on inflammation. *Nat Rev Immunol* 2010; 10(10): 712–23.
43. Gaudet AD, Popovich PG. Extracellular matrix regulation of inflammation in the healthy and injured spinal cord. *Exp Neurol* 2014; 258: 24–34.
44. Vezzani A, Baram TZ. New Roles for Interleukin-1 Beta in the Mechanisms of Epilepsy. *Epilepsy Curr* 2007; 7(2): 45–50.
45. Choi J, Nordli DR, Alden TD, Dipatri A, Laux L, Kelley K, et al. Cellular injury and neuroinflammation in children with chronic intractable epilepsy. *J Neuroinflammation* 2009; 6(1): 38.
46. Viviani B, Gardoni F, Marinovich M. Cytokines and neuronal ion channels in health and disease. *Int Rev Neurobiol* 2009; 82: 247–63.
47. Wetherington J, Serrano G, Dingledine R. Astrocytes in the epileptic brain. *Neuron* 2008; 58(2): 168–78.
48. Friedman A, Kaufner D, Heinemann U. Blood-brain barrier breakdown-inducing astrocytic transformation: Novel targets for the prevention of epilepsy. *Epilepsy Res* 2009; 85(2–3): 142–9.
49. Puttachary S, Sharma S, Stark S, Thippeswamy T. Seizure-induced oxidative stress in temporal lobe epilepsy. *Biomed Res Int* 2015; 2015: 745613.
50. Mendez-Armenta M, Nava-Ruiz C, Juarez-Rebollar D, Rodriguez-Martinez E, Gomez PY. Oxidative stress associated with neuronal apoptosis in experimental models of epilepsy. *Oxid Med Cell Longevity* 2014; 2014: 293689.
51. Pecorelli A, Natrella F, Belmonte G, Miracco C, Cervellati F, Ciccoli L, et al. NADPH oxidase activation and 4-hydroxy-2-nonenal/aquaporin-4 adducts as possible new players in oxidative neuronal damage presents in drug-resistant epilepsy. *Biochim Biophys Acta* 2015; 1852(3): 507–19.
52. Sudha K, Rao AV, Rao A. Oxidative stress and antioxidants in epilepsy. *Clin Chim Acta* 2001; 303(1–2): 19–24.
53. Warner TA, Kang JQ, Kennard JA, Harrison FE. Low brain ascorbic acid increases susceptibility to seizures in mouse models of decreased brain ascorbic acid transport and Alzheimer's disease. *Epilepsy Res* 2015; 110: 20–5.
54. Mladenović D, Djuric D, Petronijević N, Radosavljević T, Radonjić N, Matić D, et al. The correlation between lipid peroxidation in different brain regions and the severity of lindane-induced seizures in rats. *Mol Cell Biochem* 2010; 333(1–2): 243–50.
55. Ambrogini P, Minelli A, Galati C, Betti M, Lattanzi D, Ciffolilli S, et al. Post-seizure α -tocopherol treatment decreases neuroinflammation and neuronal degeneration induced by status epilepticus in rat hippocampus. *Mol Neurobiol* 2014; 50(1): 246–56.
56. Tomé Ada R, Feitosa CM, Freitas RM. Neuronal damage and memory deficits after seizures are reversed by ascorbic acid. *Arq Neuropsiquiatr* 2010; 68(4): 579–85.
57. Hrnčić D, Rašić-Marković A, Leković J, Krstić D, Čolović M, Macut D, et al. Exercise decreases susceptibility to homocysteine seizures: The role of oxidative stress. *Int J Sports Med* 2014; 35(7): 544–50.
58. Simard AR, Rivest S. Do pathogen exposure and innate immunity cause brain diseases? *Neurol Res* 2005; 27(7): 717–25.
59. Vezzani A, Aronica E, Mazarati A, Pittman QJ. Epilepsy and brain inflammation. *Exp Neurol* 2013; 244: 11–21.
60. Sayyah M, Javad-Pour M, Ghazi-Khansari M. The bacterial endotoxin lipopolysaccharide enhances seizure susceptibility in mice: Involvement of proinflammatory factors: nitric oxide and prostaglandins. *Neurosci* 2003; 122(4): 1073–80.

61. Balter-Seri J, Yubas Y, Weizman A, Nofech-Mozes Y, Kamin-sky E, Ashkenazi S. Role of nitric oxide in the enhancement of pentylenetetrazole-induced seizures caused by *Shigella dysenteriae*. *Infect. Immun.* 1999; 67(12): 6364–8.
62. Yubas Y, Weizman A, Vanichkin A, Ashkenazi S. Involvement of prostaglandins in an animal model of *Shigella*-related seizures. *J Neuroimmunol* 2005; 168(1–2): 34–9.
63. Rao RS, Medhi B, Saikia UN, Arora SK, Toor JS, Khanduja KL, et al. Experimentally induced various inflammatory models and seizure: Understanding the role of cytokine in rat. *Eur Neuropsychopharmacol* 2008; 18(10): 760–7.
64. Riazi K, Honar H, Homayoun H, Demehri S, Bahadori M, Dehpour AR. Intestinal inflammation alters the susceptibility to pentylenetetrazole-induced seizure in mice. *J. Gastroenterol. Hepatol* 2004; 19(3): 270–7.
65. Hrnčić D, Rasić-Marković A, Krstić D, Macut D, Djurić D, Stanojlović O. The role of nitric oxide in homocysteine thiolactone-induced seizures in adult rats. *Cell Mol Neurobiol* 2010; 30(2): 219–31.
66. Murta V, Farias MI, Pitossi FJ, Ferrari CC. Chronic systemic IL-1 β exacerbates central neuroinflammation independently of the blood-brain barrier integrity. *J Neuroimmunol* 2015; 278: 30–43.
67. D'Mello C, Le T, Swain MG. Cerebral microglia recruit monocytes into the brain in response to tumor necrosis factor α signaling during peripheral organ inflammation. *J Neurosci* 2009; 29(7): 2089–102.
68. Silverman HA, Dancho M, Regnier-Golanov A, Nasim M, Ochani M, Olofsson PS, et al. Brain Region-specific Alterations in the Gene Expression of Cytokines, Immune Cell Markers and Cholinergic System Components During Peripheral Endotoxin-induced Inflammation. *Mol Med* 2015; 11(20): 601–11.
69. Turrin NP, Gayle D, Ilyin SE, Flynn MC, Langhans W, Schwartz GJ, et al. Pro-inflammatory and anti-inflammatory cytokine mRNA induction in the periphery and brain following intraperitoneal administration of bacterial lipopolysaccharide. *Brain Res Bull* 2001; 54(4): 443–53.
70. Ho YH, Lin YT, Wu CW, Chao YM, Chang AY, Chan JY. Peripheral inflammation increases seizure susceptibility via the induction of neuroinflammation and oxidative stress in the hippocampus. *J Biomed Sci* 2015; 22: 46.
71. Wu KL, Chan SH, Chan JY. Neuroinflammation and oxidative stress in rostral ventrolateral medulla contribute to neurogenic hypertension induced by systemic inflammation. *J Neuroinflamm* 2012; 9: 212.
72. Rojas A, Jiang J, Ganesh T, Yang M, Lelutiu N, Gueorguieva P, et al. Cyclooxygenase-2 in epilepsy. *Epilepsia* 2014; 55(1): 17–25.
73. Takei S, Hasegawa-Ishii S, Uekawa A, Chiba Y, Umegaki H, Hosokawa M, et al. Immunohistochemical demonstration of increased prostaglandin F(2) α levels in the rat hippocampus following kainic acid-induced seizures. *Neurosci* 2012; 218: 295–304.
74. Eun BL, Abraham J, Mlsna L, Kim MJ, Koh S. Lipopolysaccharide potentiates hyperthermia-induced seizures. *Brain Behav* 2015; 5(8): e00348.

Received on September 20, 2016.

Revised on November 14, 2016.

Accepted on November 21, 2016.

Online First December, 2016.